



## Fin whale (*Balaenoptera physalus*) mitogenomics: A cautionary tale of defining sub-species from mitochondrial sequence monophyly

Cabrera, Andrea A.; Hoekendijk, Jeroen P. A.; Aguilar, Alex; Barco, Susan G.; Berrow, Simon; Bloch, Dorete; Borrell, Asunción; Cunha, Haydée A.; Dalla Rosa, Luciano; Dias, Carolina P.

Total number of authors:  
32

Published in:  
Molecular Phylogenetics and Evolution

Link to article, DOI:  
[10.1016/j.ympev.2019.02.003](https://doi.org/10.1016/j.ympev.2019.02.003)

Publication date:  
2019

Document Version  
Publisher's PDF, also known as Version of record

[Link back to DTU Orbit](#)

### Citation (APA):

Cabrera, A. A., Hoekendijk, J. P. A., Aguilar, A., Barco, S. G., Berrow, S., Bloch, D., Borrell, A., Cunha, H. A., Dalla Rosa, L., Dias, C. P., Gauffier, P., Hao, W., Landry, S., Larsen, F., Martín, V., Mizroch, S., Oosting, T., Øien, N., Pampoulie, C., ... Bérubé, M. (2019). Fin whale (*Balaenoptera physalus*) mitogenomics: A cautionary tale of defining sub-species from mitochondrial sequence monophyly. *Molecular Phylogenetics and Evolution*, 135, 86-97. <https://doi.org/10.1016/j.ympev.2019.02.003>

---

### General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.



## Fin whale (*Balaenoptera physalus*) mitogenomics: A cautionary tale of defining sub-species from mitochondrial sequence monophyly

Andrea A. Cabrera<sup>a,1,\*</sup>, Jeroen P.A. Hoekendijk<sup>a,1,4</sup>, Alex Aguilar<sup>b</sup>, Susan G. Barco<sup>c</sup>, Simon Berrow<sup>d,e</sup>, Dorete Bloch<sup>f,3</sup>, Asunción Borrell<sup>b</sup>, Haydée A. Cunha<sup>g</sup>, Luciano Dalla Rosa<sup>h</sup>, Carolina P. Dias<sup>g</sup>, Pauline Gauffier<sup>i</sup>, Wensi Hao<sup>a</sup>, Scott Landry<sup>j</sup>, Finn Larsen<sup>k,1</sup>, Vidal Martín<sup>m</sup>, Sally Mizroch<sup>n,5</sup>, Tom Oosting<sup>a</sup>, Nils Øien<sup>o</sup>, Christophe Pampoulie<sup>p</sup>, Simone Panigada<sup>q</sup>, Rui Prieto<sup>r</sup>, Christian Ramp<sup>s</sup>, Vania Rivera-Léon<sup>a</sup>, Jooke Robbins<sup>j</sup>, Conor Ryan<sup>d,e,t</sup>, Elena Schall<sup>a</sup>, Richard Sears<sup>s</sup>, Mónica A. Silva<sup>r,u</sup>, Jorge Urbán<sup>v</sup>, Frederick W. Wenzel<sup>w</sup>, Per J. Palsbøll<sup>a,j,2</sup>, Martine Bérubé<sup>a,j,2</sup>

<sup>a</sup> Marine Evolution and Conservation, Groningen Institute for Evolutionary Life Sciences, University of Groningen, Nijenborgh 7, 9747 AG Groningen, the Netherlands

<sup>b</sup> Department of Evolutionary Biology, Ecology and Environmental Sciences and Institute of Biodiversity Research (IRBio), Faculty of Biology, Universitat de Barcelona, Diagonal 643, 08028 Barcelona, Spain

<sup>c</sup> Virginia Aquarium and Marine Science Center Foundation, 717 General Booth Blvd, Virginia Beach, VA 23451, USA

<sup>d</sup> Marine and Freshwater Research Centre, Galway-Mayo Institute of Technology, Galway, Ireland

<sup>e</sup> Irish Whale and Dolphin Group, Merchant's Quay, Kilrush, County Clare V15 E762, Ireland

<sup>f</sup> Museum of National History, FO-100 Tórshavn, Faroe Islands

<sup>g</sup> Laboratório de Mamíferos Aquáticos e Bioindicadores, Faculdade de Oceanografia, Universidade do Estado do Rio de Janeiro, Rua São Francisco Xavier 524, Maracanã, 20550-013, Rio de Janeiro, Rio de Janeiro, Brazil

<sup>h</sup> Laboratório de Ecologia e Conservação da Megafauna Marinha, Instituto de Oceanografia, Universidade Federal do Rio Grande, Av. Itália km 8, Campus Carreiros, Rio Grande RS 96203-000, Rio Grande do Sul, Brazil

<sup>i</sup> Conservation, Information and Research on Cetaceans, Cabeza de Manzaneda 3, Pelayo, Algeciras 11390, Cádiz, Spain

<sup>j</sup> Center for Coastal Studies, 5 Holway Avenue, Provincetown, MA 02657, United States

<sup>k</sup> Greenland Institute of Natural Resources, c/o Box 570, Kivioq 2, 3900 Nuuk, Greenland

<sup>l</sup> Institute of Aquatic Resources, Technical University of Denmark, Kemitorvet, 2800 Kgs. Lyngby, Denmark

<sup>m</sup> Society for Study of Cetaceans in the Canary Archipelago, Avenida Coll 6, (Casa de Los Arroyo) 35500 Arrecife de, Lanzarote – Islas Canarias, Spain

<sup>n</sup> National Marine Mammal Laboratory, Alaska Fisheries Science Center/NOAA, 7600 Sand Point Way N.E., Seattle, WA 98115, United States

<sup>o</sup> Marine Mammals Research Group, Institute of Marine Research, C. Sundtsgate 64, P. O. Box 1870 Nordnes, Bergen N-5817, Norway

<sup>p</sup> Marine and Freshwater Research Institute, Skúlagata 4, 121 Reykjavík, Iceland

<sup>q</sup> Tethys Research Institute, c/o Acquario Civico, Viale G.B. Gadio 2, 20121 Milano, Italy

<sup>r</sup> Marine and Environmental Sciences Centre and Centre of the Institute of Marine Research of the University of the Azores, R. Frederico Machado 4, 9901-862 Horta, Portugal

<sup>s</sup> Mingan Island Cetacean Study Inc., 285 Green St, Saint-Lambert, Québec J4P 1T3, Canada

<sup>t</sup> Song of the Whale Research Team, Marine Conservation Research International, 94 High Street, Kelvedon, Essex CO5 9AA, United Kingdom

<sup>u</sup> Biology Department, Woods Hole Oceanographic Institution, Woods Hole, MA 02543, United States

<sup>v</sup> Departamento de Ciencias Marinas y Costeras, Universidad Autónoma de Baja California Sur, La Paz 23080, B.C.S., Mexico

<sup>w</sup> Woods Hole MA Laboratory, Northeast Fisheries Science Center NMFS/NOAA, 166 Water Street, Woods Hole, MA 02543, United States

### ARTICLE INFO

#### Keywords:

Fin whale

*Balaenoptera physalus*

### ABSTRACT

The advent of massive parallel sequencing technologies has resulted in an increase of studies based upon complete mitochondrial genome DNA sequences that revisit the taxonomic status within and among species. Spatially distinct monophyly in such mitogenomic genealogies, i.e., the sharing of a recent common ancestor

\* Corresponding author.

E-mail addresses: [andrea\\_ca\\_gt@yahoo.com](mailto:andrea_ca_gt@yahoo.com) (A.A. Cabrera), [palsboll@gmail.com](mailto:palsboll@gmail.com) (P.J. Palsbøll), [m.berube@rug.nl](mailto:m.berube@rug.nl) (M. Bérubé).

<sup>1</sup> Equally contributing authors.

<sup>2</sup> Senior corresponding authors.

<sup>3</sup> Deceased.

<sup>4</sup> Current address: NIOZ Royal Netherlands Institute for Sea Research, Department of Coastal Systems (COS), and Utrecht University, P.O. Box 59, 1790 AB Den Burg, Texel, the Netherlands.

<sup>5</sup> Current address: Blue Sea Research, PO Box 15805, Seattle, WA 98115, USA.

<https://doi.org/10.1016/j.ympev.2019.02.003>

Received 30 October 2018; Received in revised form 30 January 2019; Accepted 4 February 2019

Available online 13 February 2019

1055-7903/ © 2019 The Authors. Published by Elsevier Inc. This is an open access article under the CC BY-NC-ND license

(<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

North Atlantic Ocean  
Subspecies  
Mitochondrial genome

among con-specific samples collected in the same region has been viewed as evidence for subspecies. Several recent studies in cetaceans have employed this criterion to suggest subsequent taxonomic revisions. We reason that employing intra-specific, spatially distinct monophyly at non-recombining, clonally inherited genomes is an unsatisfactory criterion for defining subspecies based upon theoretical (genetic drift) and practical (sampling effort) arguments. This point was illustrated by a re-analysis of a global mitogenomic assessment of fin whales, *Balaenoptera physalus* spp., published by Archer et al. (2013), which proposed to further subdivide the Northern Hemisphere fin whale subspecies, *B. p. physalus*. The proposed revision was based upon the detection of spatially distinct monophyly among North Atlantic and North Pacific fin whales in a genealogy based upon complete mitochondrial genome DNA sequences. The extended analysis conducted in this study (1676 mitochondrial control region, 162 complete mitochondrial genome DNA sequences and 20 microsatellite loci genotyped in 380 samples) revealed that the apparent monophyly among North Atlantic fin whales reported by Archer et al. (2013) to be due to low sample sizes. In conclusion, defining sub-species from monophyly (i.e., the absence of para- or polyphyly) can lead to erroneous conclusions due to relatively “trivial” aspects, such as sampling. Basic population genetic processes (i.e., genetic drift and migration) also affect the time to the most recent common ancestor and hence the probability that individuals in a sample are monophyletic.

## 1. Introduction

Genealogies estimated from mitochondrial DNA (mtDNA) sequences have been employed towards resolving inter- and intraspecific taxonomic relationships for more than three decades (Avice, 1989; Ball and Avice, 1992; Burbrink et al., 2000). Taxonomic assessments aimed below the nominal species level usually focus on the spatial distinctiveness of monophyletic clades in genealogies estimated from mtDNA sequences, i.e., the presence of phylogeographic structure (Avice et al., 1979, 1987; Ball and Avice, 1992). The presence of spatially confined monophyletic mitochondrial clades has typically been inferred as evidence for a high level of reproductive isolation and consequently some degree of evolutionary distinctiveness. Evolutionary significant units (ESUs) serve as an illustrative example (Ryder, 1986; Bernatchez, 1995). ESUs are generally viewed as distinct components of intraspecific genetic diversity (Ryder, 1986; Bernatchez, 1995). Moritz (1994) proposed that ESUs were defined by their reciprocal monophyly, specifically in genealogies estimated from mtDNA sequences. In addition, allele frequencies at nuclear loci should be “significantly” different between reciprocally monophyletic mtDNA sequence clades. When monophyly in a mtDNA genealogy is employed as the defining criterion, a key question becomes whether this level of phylogeographic structure always equates to isolation and evolutionary distinctiveness. In other words, does the *absence* of monophyly implies a recent common ancestry and evolutionary indistinctiveness? Paetkau (1999) pointed to the fact that the effective population size and the time to the most recent common ancestor (TMRCA) are positively correlated. This fundamental relationship implies that isolated populations at low effective population sizes will become monophyletic faster than populations with large effective population sizes. This difference has immediate ramifications in those cases when mtDNA monophyly is employed as the main, or sole, criterion in defining ESUs (e.g., Banguera-Hinestroza et al., 2002; Archer et al., 2013).

Similarly, low sampling effort may lead to erroneous conclusions when defining ESUs from spatial monophyly in mitogenomic genealogies. Apparent monophyly could be simply a product of insufficient sampling, i.e., an insufficient number of specimens to capture all mtDNA clades (Funk and Omland, 2003). Intraspecific genealogies inferred from mtDNA sequences often contain multiple well-supported clades. However, the relative proportions of such clades typically vary across space. Consequently, insufficient sampling in all, or some regions, may result in failure to sample DNA sequences belonging to uncommon clades, erroneously leading to the conclusion of monophyly (Funk and Omland, 2003).

Initially most phylogeographic studies were based solely upon genealogies inferred from mtDNA sequence variation (Ball and Avice, 1992; Burbrink et al., 2000; Pons et al., 2006). The mtDNA genome (mitogenome) was viewed as especially suitable for this kind of assessments due to its haploid, often maternal and clonal inheritance,

which alleviates potential issues in inferring genealogies from recombining nuclear loci. However, several studies have demonstrated that inferring intraspecific isolation from mtDNA sequences only, could be misleading, ironically because of the maternal inheritance, which prevented detection of male mediated gene flow (Prager et al., 1993; Palumbi and Baker, 1994). Consequently, many studies have since complemented mtDNA sequences with nuclear, biparentally-inherited DNA sequences in phylogeographic analyses aimed at assessing evolutionary distinctiveness, such as ESUs as proposed by Moritz (1994).

The relatively recent development of affordable massive parallel sequencing technologies (Funk et al., 2012) has led to a resurgence in phylogeographic studies based solely on mtDNA sequences, albeit of the complete mitogenome as opposed to a few hundreds of base pairs (Morin et al., 2004, 2010; Archer et al., 2013; Meng et al., 2013). A search in Web of Science™ (Clarivate Analytics Inc.) revealed that only 14 out of 100 publications assessing phylogeographic structure or intraspecific taxonomic revisions, complemented complete mitogenome sequence data with data from nuclear loci (see [Supplementary Materials](#)). The sample sizes in studies based on complete mitogenome sequences in non-model species (Morin et al., 2004, 2010; Archer et al., 2013; Meng et al., 2013) remains considerably lower compared to contemporaneous studies based upon Sanger (1981) DNA sequencing of smaller mtDNA regions and nuclear loci (Pastene et al., 2007; Halbert et al., 2013; Jackson et al., 2014). These two aspects, relying solely on mitogenome sequence data (Zachos et al., 2013) and low sample sizes, implies that the detection of monophyly is prone to the caveats that haunted earlier, similar studies based upon shorter mtDNA sequences, such as the mtDNA control region (CR). Studies based upon complete mitogenome sequences typically yield very high support for the basal nodes, leading to the impression of high accuracy. However, high accuracy in a single-locus genealogy does not necessarily imply that the genealogy accurately reflects the population/subspecies history as has been pointed out by numerous authors in the past (Pamilo and Nei, 1988; Page and Charleston, 1997; Leaché, 2009).

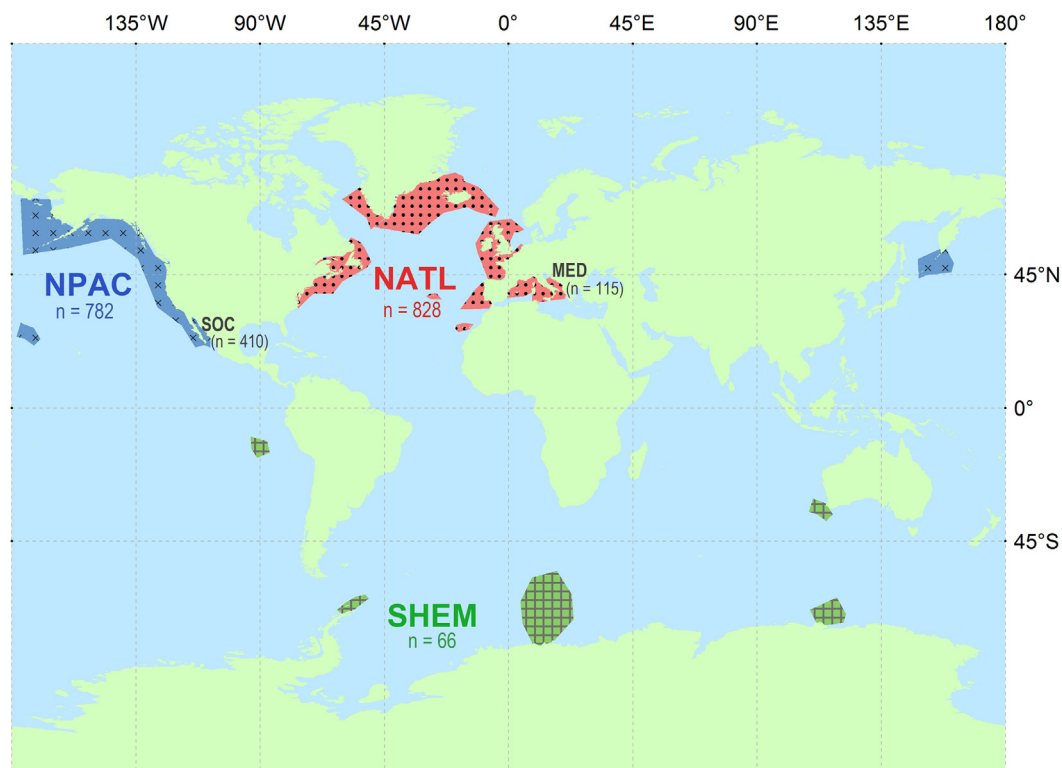
A case in point is *Cetacea* (whales, dolphins and porpoises), a group of highly derived mammals, which has recently been subjected to several re-assessment of species/subspecies status based upon the estimation of intraspecific genealogies from complete mitogenome sequences (Morin et al., 2010; Vilstrup et al., 2011; Archer et al., 2013). The large body sizes, wide ranges and limited availability of osteological specimens in most cetacean species has made it difficult to apply traditional, non-molecular approaches to define intra-specific taxonomic entities and explains the popularity of molecular-based taxonomic assessments in cetaceans. Most baleen whale (*Mysticeti*) species have global distributions and migrate seasonally between low latitude winter breeding grounds and high latitude summer feeding grounds (Ingebrigtsen, 1929; Dawbin, 1966; Jonsgård, 1966; Katona and Whitehead, 1981). As a result, most baleen whale populations roam across entire ocean basins making it challenging to delineate intra-

specific evolutionary units. Two aspects are generally assumed, *a priori*, to confine baleen whale distributions and restrict gene flow. The anti-tropical distribution of most baleen whale species presumably acts as a reproductive barrier between the two hemispheres, despite the (proximate) low latitude locations of winter breeding grounds, because the breeding season for each hemisphere is separated by half a year (Davis et al., 1998). In addition, most ocean basins are separated by the continents, which prevent inter-oceanic dispersal as well. Consequently, it is generally assumed that gene flow between con-specific baleen whale populations in different ocean basins is very limited (Valsecchi et al., 1997; Bérubé et al., 1998; Pastene et al., 2007; Jackson et al., 2014). Accordingly, current, recognized baleen whale species and subspecies designations typically correspond to ocean basins or hemispheres. For instance, the right whales are comprised of *Eubalaena glacialis*, in the North Atlantic; *E. australis*, in the Southern Hemisphere; and *E. japonica*, in the North Pacific (Rice, 1998; Rosenbaum et al., 2000). Similarly Northern Hemisphere blue whales, *Balaenoptera musculus*, are classified as *B. m. musculus* and Southern Hemisphere blue whales as *B. m. intermedia*, in addition to the pygmy blue whale, *B. m. brevicauda* (Rice, 1998).

The fin whale, *Balaenoptera physalus* spp. (Linnaeus, 1758), is a common and globally distributed baleen whale (Gambell, 1985). Fin whales in the Northern Hemisphere are classified as belonging to the subspecies *B. p. physalus* and fin whales in the Southern Hemisphere to *B. p. quoyi* (Fischer, 1829). The fin whale subspecies designations were based upon differences in the vertebrate characteristics (Lönnerberg, 1931) as well as traits correlated with body size (Tomilin 1946 cited by Rice, 1998). Employing this classification, North Pacific and North Atlantic fin whales both belong to the same subspecies, despite the observation that gene flow between the two ocean basins is unlikely, at least since the rise of the Panama Isthmus approximately 3.5 million years ago (Coates et al., 1992). Recently, Archer et al. (2013) employed

complete mitogenome sequences from North Atlantic, North Pacific and Southern Hemisphere fin whale specimens to assess the current subspecies status of Northern Hemisphere fin whales. Archer et al. (2013) concluded that North Atlantic and some North Pacific fin whales constituted separate subspecies. This conclusion was based upon the observation of a single monophyletic clade that contained all North Atlantic specimens (a sample of 14 specimens), and the presence of several monophyletic clades containing solely North Pacific specimens (Fig. 3a).

The results of Archer et al.'s (2013) mitogenomic analysis appeared at odds with previous phylogeographic assessments by Bérubé et al. (1998, 2002). Bérubé and colleagues based their assessments upon DNA sequences from the highly variable mtDNA CR. Their study identified two mtDNA CR haplotypes in North Atlantic specimens that clustered together with mtDNA CR haplotypes identified among North Pacific specimens. Bérubé and co-workers inferred this result as evidence for recent gene flow between the North Atlantic and North Pacific (Bérubé et al., 1998, 2002) likely in a stepping stone manner via the Southern Ocean. In order to resolve the discrepancy between the above-mentioned studies and the support for the taxonomical revision proposed by Archer et al. (2013), this study extended the sample size of North Atlantic Ocean (including the Mediterranean Sea) fin whales from the 34 mtDNA CR sequences analyzed by Archer et al. (2013) to a total of 828 mtDNA CR sequences. The complete mitogenome was sequenced in a subset ( $n = 6$ ) of North Atlantic specimens, including mtDNA CR haplotypes that clustered with mtDNA CR haplotypes detected in specimens sampled outside the North Atlantic. In addition, 20 micro-satellite loci were genotyped in 358 specimens from the North Atlantic, North Pacific and Southern Hemisphere. The re-estimation of the genealogy based upon the complete mitogenome sequences from this study showed all ocean basins to be polyphyletic. In other words, these results did not support the current nor the proposed subspecies if monophyly



**Fig. 1.** Sampling location from the North Atlantic, North Pacific and Southern Hemisphere fin whales. Notes: Approximate sampling locations in the North Atlantic (NATL, red), North Pacific (NPAC, blue) and the Southern Hemisphere (SHEM, green). The numbers represent the total number of mitochondrial control region DNA sequences from each region. Numbers in black denote the sample sizes from the Mediterranean Sea (MED, included in NATL) and the Sea of Cortez (SOC, included in NPAC). (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)



in a genealogy estimated from mtDNA sequences is employed as the sole or main defining criterion. The basal topology of the genealogies estimated from the mitogenome and mtDNA CR sequences were qualitatively similar as expected given that the mitogenome represents one linked locus. The assignment test based on the genotype of 20 microsatellite loci revealed that the North Atlantic specimens from the two different mitochondrial DNA clades all belonged to the same North Atlantic gene pool.

The findings of this study highlight the implications of insufficient sampling when attempting to identify monophyletic clades from mtDNA sequences. However, the results did not negate the possibility that fin whales from different ocean basins could potentially represent different subspecies, although the analysis from this study revealed recent gene flow between fin whales from different ocean basins and hemispheres.

More generally, employing monophyly in genealogies based upon DNA sequences from non-recombining genomes to classify subspecies ignores fundamental population genetic processes as well as key practical issues. These caveats make the approach less valid than its current widespread use suggests. Although these caveats have been highlighted earlier (Paetkau, 1999; Funk and Omland, 2003), the approach has nevertheless regained momentum given the ease of applying massive parallel sequencing technologies to uniparentally inherited, non-recombining genomes, such as the mitogenome.

## 2. Materials and methods

### 2.1. Sample collection

Tissue samples were obtained from fin whales in the North Atlantic Ocean basin and the Mediterranean Sea (henceforth referred to collectively as the North Atlantic); the North Pacific Ocean basin and the Sea of Cortez (henceforth referred to collectively as the North Pacific) as well as the Southern Hemisphere between 1982 and 2014 (Fig. 1). Most tissue samples were collected as skin biopsies from free-ranging fin whales as described by Palsbøll et al. (1991). The tissue samples originating from Iceland and Spain were collected during whaling operations prior to the international moratorium on commercial whaling. Some samples collected in Greenland originated from local subsistence whaling and some samples collected in US waters from dead, beached individuals. All samples were collected in agreement with national and international regulations. Samples were preserved in 5 M NaCl with 20% dimethyl sulfoxide and stored at –20 degrees Celsius (Amos and Hoelzel, 1991).

### 2.2. Mitochondrial DNA sequence data

The mtDNA sequence data were either generated during this study or obtained from the study by Archer et al. (2013) who deposited the data in the Dryad repository (<https://doi.org/10.5061/dryad.084g8>). The experimental methods used to generate the published data were described by Archer et al. (2013).

The DNA sequence data generated for this study were obtained in the following manner. Total-cell DNA was extracted from tissue samples either by phenol/chloroform extraction as described by Sambrook and Russell (2001) or using the Qiagen DNEasy™ Blood and Tissue Kit columns (QIAGEN Inc., Valencia, CA, USA) following the manufacturer's instructions. Samples were sexed using the ZFY/ZFX multiplexing system as described by Bérubé and Palsbøll (1996b); Bérubé and Palsbøll (1996a). MtDNA CR DNA sequencing was performed as previously described either by (i) Palsbøll et al. (1995), but replacing the original reverse primer with BP16071, (Drouot et al., 2004); or (ii) Bérubé et al. (2002). The complete mitogenome was sequenced in eight selected specimens. These specimens were selected from genealogy estimated from all mtDNA CR haplotypes. Six specimens were selected among the 33 North Atlantic specimens with mtDNA CR haplotypes

that clustered with specimens sampled in other ocean basins. The remaining two specimens were selected among specimens from the North Atlantic and North Pacific Ocean, respectively, with mtDNA CR haplotypes that clustered with other specimens sampled in the same ocean within monophyletic clades. A total of 35 nested primer pairs were employed (Supplementary Materials, Table S1) to amplify and determine the DNA sequence of the complete fin whale mitogenome from partially overlapping ~500 base pair (bp) fragments. PCR (Mullis and Faloona, 1987) amplifications and DNA sequencing were performed under conditions identical to those described above for the mtDNA CR sequencing albeit at different annealing temperatures (Supplementary Materials, Table S1). Phred quality scores were employed to assess the sequence quality and discard low-quality sequences (phred score below 20). The quality of the previously published data is described by Archer et al. (2013).

### 2.3. Microsatellite genotyping

The genotype was determined at 20 diploid, autosomal microsatellite loci in samples from four different regions; the North Atlantic (including the Mediterranean Sea) (n = 288), the Eastern North Pacific (n = 25), the Sea of Cortez (n = 46) and the Southern Hemisphere (i.e., the Antarctic, n = 21). The specific microsatellite loci were: AC087, CA234 (Bérubé et al., 2005), EV00, EV037, EV094 (Valsecchi and Amos, 1996), GATA028, GATA098, GATA417 (Palsbøll et al., 1997), GATA25072, GATA43950, GATA5947654, GATA6063318, GATA91083 (Bérubé et al., in prep), GT011 (Bérubé et al., 1998) GT023, GT211, GT271, GT310, GT575, (Bérubé et al., 2000) and TAA023 (Palsbøll et al., 1997) with tetra, tri- or dimer repeat motifs (Supplementary Materials, Table S2). Individual PCR amplifications were performed in 10 µL volumes, each containing ~2–10 ng of extracted DNA, 0.2 µM of each oligo-nucleotide primer (Supplementary Materials, Table S2) and 1X final QIAGEN Microsatellite PCR Multiplex Mix™ (Qiagen Inc.). Thermo-cycling was carried out on a MJ Research PTC-100™ Thermal Cycler (BioRad Inc.). The PCR amplification consisted of an initial step of five minutes at 95 degrees Celsius, followed by 35 cycles; each of 30 s at 95 degrees Celsius, 90 s at 57 degrees Celsius and 30 s at 72 degrees Celsius. The final step was 10 min at 68 degrees Celsius. PCR reactions were diluted 60 times with MilliQ water and then 1 µL of diluted PCR reaction was added to 9 µL of GeneScan-500™ ROX (Applied Biosystems Inc.) and deionized formamide (GeneScan-500™ ROX 1 µL: 70 µL) prior to electrophoresis on an ABI 3730 Genetic Analyzer™ (Applied Biosystems Inc.). The length of each amplification product was determined using GeneMapper™ ver. 4.0 (Applied Biosystems Inc.).

### 2.4. Assembly and analysis of the mitochondrial DNA sequences

MtDNA sequences were aligned and assembled against the fin whale mitogenome sequence deposited in GenBank™ (accession # NC001321) by Árnason et al. (1991) using SEQMAN™ (ver. 5.05, DNASTAR Inc.) with default parameter settings. All DNA sequences were trimmed to equal length, i.e., 16,423 and 285 bp for the mitogenome and mtDNA CR DNA sequences, respectively.

*Estimation of mtDNA haplotype genealogies and divergence times:* The genealogies of the mtDNA CR and complete mitogenome haplotypes as well as divergence times, were estimated employing the software BEAST (ver. 1.8.2, Drummond and Rambaut, 2007; Drummond et al., 2012) largely following the approach by Archer et al. (2013). However, in contrast to Archer et al. (2013), only a single copy of each haplotype from each ocean basin (both complete and CR mtDNA sequences) was included in each data set. Insertion and deletions were coded as a fifth character. Genealogies were rooted with the homolog DNA sequence from humpback whale, *Megaptera novaeangliae*, (GenBank™ accession #NC006927, Sasaki et al. (2005)) using the alignment reported by Archer et al. (2013). The most probable nucleotide

mutation model and associated parameter values (e.g., the transition:transversion ratio, the proportion of invariable sites and the gamma distribution) was determined using software JMODELTEST (ver. 2, Darriba et al., 2012) and selected using the Bayesian information criterion. The HKY + I + G substitution model (Hasegawa et al., 1985) with four substitution categories was selected. A strict molecular clock with a uniform prior distribution and rates between  $1 \times 10^{-5}$  and  $1 \times 10^{-2}$  substitutions per site per million years was assumed. Similar to Archer et al. (2013), the Yule speciation model was employed and the tree topology and branch lengths were initialized with the unweighted pair group method and the arithmetic mean. The TMRCA between the fin whale and humpback whale (Sasaki et al., 2005) was employed as the prior for the root of the genealogy. The range was set with a normal distribution at 2.8–15.8 million years. The genealogy and the posterior distribution of the divergence times were estimated using Monte Carlo Markov chains (MCMC) sampling. Samples were drawn at every 1000th step from a total of  $2 \times 10^7$  steps of which the first 10% was discarded as burn-in. Convergence to stationarity and mixing were evaluated with TRACER (ver. 1.5, Rambaut and Drummond, 2007). The consensus genealogy as well as the posterior probability for major nodes and divergence times were obtained using TREEANNOTATOR (ver. 1.8.3), as implemented in BEAST.

**Estimation of genetic diversity and immigration rates:** The software MIGRATE-N (ver. 3.6.6, Beerli and Felsenstein, 1999, 2001) was employed to estimate the genetic diversity ( $\Theta$ ) and immigration rate scaled by the generational mutation rate ( $M$ ) per nucleotide site among the North Atlantic, North Pacific and Southern Hemisphere. The prior ranges of  $\Theta$  and  $M$  were determined from preliminary estimations with reduced sample sizes and short MCMC chains with the  $F_{ST}$ -based method as starting values. The prior ranges were subsequently adjusted according to the outcomes of these preliminary estimations, i.e.,  $\Theta$  (uniform prior, min: 0, max: 0.25,  $\delta$ : 0.025) and  $M$  (uniform prior, min: 0, max: 250.0,  $\delta$ : 25.0). Data sets above 100 DNA sequences randomly sub-sampled (without replacement) at sample sizes of 100 DNA sequences per sample partition. Due to significant levels of intra-ocean population structure in mtDNA DNA sequence variation (Bérubé et al., 1998; Palsbøll et al., 2004; Rivera-León et al., under review), samples from the Mediterranean Sea and the Sea of Cortez were excluded. The final estimates were inferred from a total of three independent estimations. Each estimation was initiated with a different random seed and comprised 100 replicates, each consisting of a single long MCMC with 10 million steps discarded as burn in followed by an additional 10 million steps, sampled at every 200th step. A static heating scheme of four chains at temperatures 1.0, 1.5, 3.0, and 1,000,000, respectively, was employed. Convergence was assessed employing the R-CRAN package CODA (ver. 0.19-1, Plummer et al., 2006). Consistency among

the three independent estimations, smooth and unimodal distribution within the prior range and an effective sample size above 100,000 for all parameter estimates were also considered as indications of convergence (Supplementary Materials, Table S3).

### 2.5. Multi-locus genotype assignments

The likelihood of multi-locus microsatellite genotypes given the observed allele frequencies in each putative source population was estimated using the probability of identity (Paetkau and Strobeck, 1994) as implemented in GENECLASS v. 2.0 (Piry et al., 2004). The null-distribution was estimated from 10,000 multi-locus genotypes drawn at random with replacement from the observed data. The type 1 error rate was set to the default value at 0.01. The observed allele frequencies in each putative source population was estimated from 266 multi-locus microsatellite genotypes for the North Atlantic (including the Mediterranean Sea), 24 for the Eastern North Pacific, 45 for the Sea of Cortez and 20 for the Southern Hemisphere (i.e., the Antarctic).

### 3. Results

The final data sets comprised 1676 fin whale mtDNA CR DNA sequences and 162 fin whale complete mitogenome sequences (Table 1). Among the final 1676 mtDNA CR DNA sequences, 428 DNA sequences were obtained from Archer et al. (2013) and 1248 were generated for this study. A total of 410 mtDNA CR sequences from the 782 North Pacific were collected in the Sea of Cortez, a population with low mtDNA sequence diversity (Bérubé et al., 2002). Among the 828 North Atlantic mtDNA CR sequences, 115 were collected in the Mediterranean Sea (Fig. 1). A total of 161 haplotypes were detected among the 1676 fin whale mtDNA CR sequences, (Table 1) and 147 haplotypes among the 162 complete mitogenome sequences of which 154 were published by Archer et al. (2013) and eight generated during this study (Table 1).

Both genealogies estimated from the complete mitogenome and mtDNA CR haplotypes published by Archer et al. (2013) identified a single monophyletic clade containing all, and only, North Atlantic specimens (denoted NA clade in Figs. 2a and 3a). In contrast, haplotypes detected in North Pacific and Southern Hemisphere specimens were polyphyletic (Figs. 2a and 3a). In contrast, the genealogies estimated from the complete mitogenome and mtDNA CR haplotypes including all data, i.e., both the data generated for this study and those published by Archer et al. (2013), partitioned the North Atlantic specimens in two major clades; one clade (denoted NA clade in Fig. 2b) comprised solely of North Atlantic specimens and another clade comprised DNA sequence haplotypes detected in specimens from the North Pacific and Southern Hemisphere, in addition to the North Atlantic

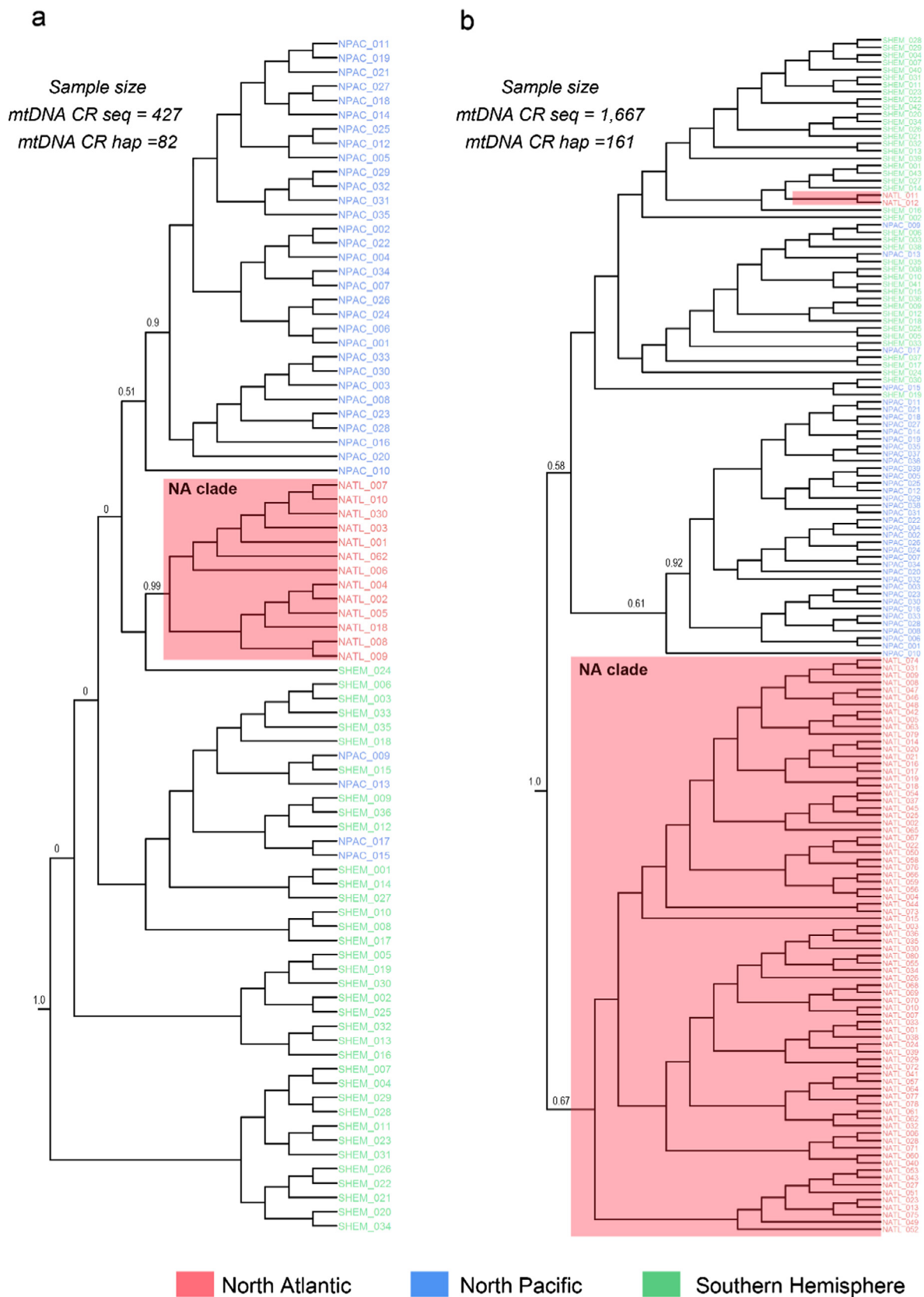
**Table 1**  
Mitochondrial DNA control region (mtDNA CR) and complete mitochondrial DNA genome (mitogenome) sequences and haplotypes per ocean basin.

	North Atlantic*		North Pacific**		Southern Hemisphere		Total	
	SEQ	HAP	SEQ	HAP	SEQ	HAP	SEQ	HAP
<b>MtDNA CR</b>								
Archer et al. (2013) <sup>a</sup>	34	13	346	35	48	36	428	83
This study	794	80	436	14	18	11	1248	104
Combined data	828	80	782	39	66	43	1676	161
<b>Complete mitogenome</b>								
Archer et al. (2013)	14	12	97	89	43	42	154	143
This study	7	4	1	1	0	0	8	5
Combined data	21	16	98	89	43	42	162	147

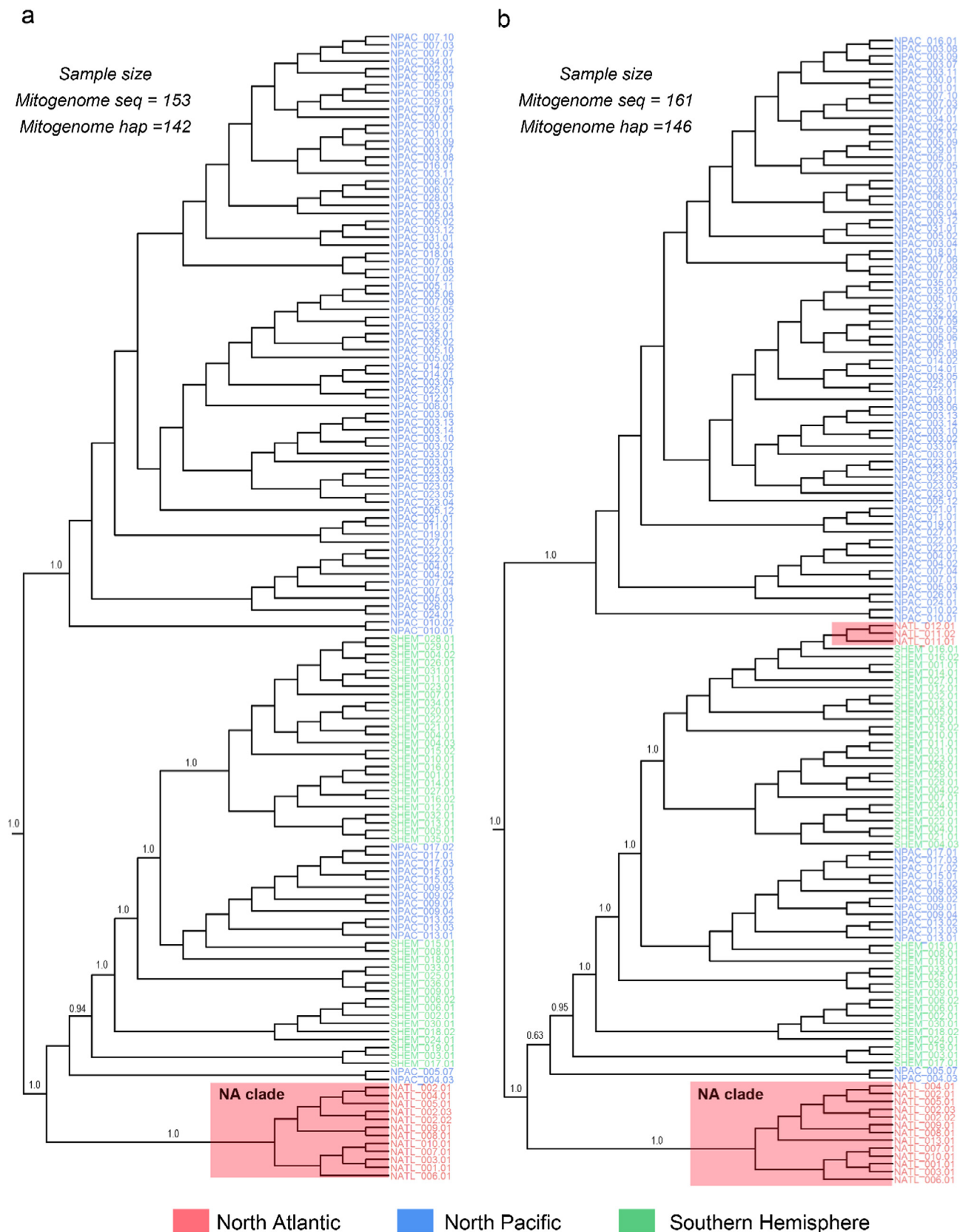
<sup>a</sup> The data was recreated from the sample information file (<http://datadryad.org/bitstream/handle/10255/dryad.48318/Bphy%20sample%20info.csv?sequence=1>) deposited by Archer et al. (2013) in the Dryad data repository. The file contains GenBank accession numbers for each sample entry (either only mtDNA CR sequence (n = 274), or the mtDNA CR sequence extracted from the complete mitogenome sequence (n = 154). SEQ: number of sequences, HAP: number of unique haplotypes.

\* North Atlantic including Mediterranean Sea.

\*\* North Pacific including Sea of Cortez.



**Fig. 2.** Bayesian genealogy estimated from North Atlantic, North Pacific and Southern Hemisphere fin whale mitochondrial control region (mtDNA CR) haplotypes. Notes: Genealogies were estimated from (a) 82 mtDNA CR haplotypes reported by Archer et al. (2013) and (b) 161 mtDNA CR haplotypes reported by Archer et al. (2013) combined with additional mtDNA CR haplotypes reported in this study. Colors represent the three ocean basins/regions: the Southern Hemisphere (green, denoted SHEMA), the North Pacific (blue, denoted NPAC) and the North Atlantic (red, denoted NATL), respectively. Numbers at basic nodes denotes the posterior probability of the specific node (only the support for basic nodes is reported). A humpback whale (*Megaptera novaeangliae*) mtDNA CR haplotype (Genbank NC\_006927) was employed to root the tree (not shown). (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)



**Fig. 3.** Bayesian genealogy estimated from North Atlantic, North Pacific and Southern Hemisphere fin whale mitochondrial genome (mitogenome) haplotypes. Notes: Genealogies were estimated from (a) 142 mitogenome haplotypes reported by Archer et al. (2013) and (b) 146 mitogenome haplotypes reported by Archer et al. (2013) combined with additional mitogenome haplotypes reported in this study. Colors represent the three ocean basins/regions: the Southern Hemisphere (green, denoted SHER), the North Pacific (blue, denoted NPAC) and the North Atlantic Ocean (red, denoted NATL), respectively. Numbers at basic nodes denotes the posterior probability of the specific node (only the support for basic nodes is reported). A humpback whale (*Megaptera novaeangliae*) mitogenome haplotype (Genbank NC\_006927) was employed to root the tree (not shown). (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)



(haplotypes NATL\_011 and NATL\_012, Fig. 2b). The genealogy estimated from the novel and previously published complete mitogenome haplotypes was similar to the genealogy inferred from the mtDNA CR sequences (Figs. 2b and 3b). In agreement with the genealogy estimated from the mtDNA CR sequences, North Atlantic specimens were partitioned into two different clades; one clade containing solely North Atlantic specimens (NA clade, Fig. 3b) and another clade containing specimens from the North Pacific, Southern Hemisphere and North Atlantic (Fig. 3b). The latter clade contained three haplotypes (haplotypes NATL\_011.01, NATL\_011.02 and NATL\_012.01, Fig. 3b) represented by all six North Atlantic specimens from which complete mitogenome DNA sequences were obtained during this study (Fig. 3b).

The TMRCA estimated from all complete mitogenome haplotypes included this study (Fig. 3b) was estimated at 1.9 million years and the 95% HPD (highest probability density) interval from 1.1 to 2.8 million years (Table 2). The divergence time of the three North Atlantic complete mitogenome haplotype, which clustered outside the NA clade (Fig. 3b) detected during this study was estimated at 0.095 million years and the 95% HPD interval from 0.04 to 0.17 million years. The TMRCA for all the complete mitogenome haplotypes detected in the North Atlantic was estimated at 0.99 million years and the 95% HPD from 0.54 to 1.4 million years (Table 2). This estimate was 0.45 million years older than that reported by Archer et al. (2013). The TMRCA for all the mtDNA CR haplotypes included in this study (Fig. 2b) was estimated at 4.3 million years and the 95% HPD interval from 1.97 to 6.8 million years. In the case of the North Atlantic fin whales, the TMRCA was estimated at 4.2 million years and the 95% HPD interval from 1.96 to 6.8 million years.

The population origin of 25 fin whale specimens (22 sampled in the North Atlantic, one in the North Pacific, one in the Sea of Cortez and one in the Southern Hemisphere) was inferred from the assignment tests based upon diploid genotypes at 20 microsatellite loci. Among the 22 fin whale specimens from the North Atlantic, 20 samples had mtDNA CR haplotypes that were assigned to clades that also contained specimens from the North Pacific and Southern Hemisphere (i.e., outside clade “NA”, specifically haplotypes NATL\_011 and NATL\_012, Fig. 2b). The remaining two North Atlantic specimens had mtDNA CR haplotypes that were assigned to the main North Atlantic clade (clade “NA”, specifically haplotypes NATL\_003 and NATL\_016, Fig. 2b). All specimens were assigned to the population in which they were sampled with probabilities  $\sim 1.0$  (Table 4).

The immigration rate (i.e.,  $N_e m$ ) estimated from the mtDNA CR sequences from the Southern Hemisphere into the North Pacific was estimated at 0.36 (95% credible interval: 0–3.41, Table 3) which is equivalent to a single immigrant per 2.8 generations. The immigration rates from the Southern Hemisphere into the North Atlantic and from the North Atlantic into the North Pacific were estimated at 0.37 (95% credible interval: 0–2.79, Table 3) and 0.0015 (95% credible interval:

0–1.56, Table 3), respectively. Lastly the immigration rate from the North Pacific into the North Atlantic was estimated at 0.0029 (95% credible interval: 0–1.97, Table 3).

#### 4. Discussion

The initial reason for undertaking this study was the discrepancies between Archer et al. (2013) findings and the earlier work published by Bérubé et al. (2002, 1998). However, there was a more general concern about the recent resurgence in mitogenomic-based studies employing monophyly to delineate intraspecific evolutionary distinct units.

Meiotic recombination facilitates the incorporation of population-wide variation into each haploid genome complement (Pamilo and Nei, 1988). Accordingly, population-specific monophyly at recombining loci requires substantial reproductive isolation during a considerable time. How long depends upon the effective population size (see Hudson and Turelli, 2003). The situation is different for a uniparentally inherited, non-recombining genome where each lineage contains only the variation of its own lineage rather than the population at large, which is why non-recombining loci are sensitive to the sampling effects illustrated in this study. This difference highlights the need for a sufficiently exhaustive sampling scheme. It appears that sampling was the cause for the monophyly of North Atlantic fin whales observed by Archer et al. (2013). Archer et al. (2013) included a total of 34 North Atlantic fin whale specimens (including Mediterranean Sea specimens) in their analysis, represented by 13 haplotypes. The extended sample in this study comprised 828 North Atlantic fin whale mtDNA CR sequences representing 80 haplotypes. Among the 828 sequences, 33 (i.e.,  $\sim 4\%$ ) had one of the two haplotypes that clustered outside the main North Atlantic clade (NA clade in Fig. 2b). The relative scarcity of North Atlantic fin whales that carry mtDNA haplotypes clustering outside the North Atlantic could be the result of recent dispersal into the North Atlantic. Consequently, North Atlantic fin whales with these mtDNA haplotype may simply be immigrants and not actually part of the North Atlantic gene pool *per se*. However, the analyses of the biparentally inherited microsatellite loci in this study suggested that these individuals were part of the North Atlantic gene pool and unlikely to originate from the North Pacific (both the Sea of Cortez and the eastern North Pacific) nor the Southern Hemisphere. The probability of all these samples' multi-locus genotypes was higher in the North Atlantic and Mediterranean Sea than in the other ocean basins (Table 4).

The divergence times among the basic nodes estimated during this study, (Table 2) were considerably longer than the estimates reported by Archer et al. (2013). These differences were mainly due to polyphyly detected in the expanded sample from the North Atlantic. As a result, the TMRCA among North Atlantic specimens in this study was much longer. Another consequence of a polyphyletic North Atlantic samples was that the intra-oceanic and global TMRCA were similar. The

**Table 2**

Estimates of time to most recent common ancestor and substitution rates obtained from the mitochondrial control region and the entire mitochondrial genome DNA sequences.

	North Atlantic*		North Pacific**		Southern Hemisphere		All three ocean basins		Substitution rate	
	TMRCA	95% HPD	TMRCA	95% HPD	TMRCA	95% HPD	TMRCA	95% HPD	mean	95% HPD
<i>Mitochondrial control region</i>										
Archer et al. (2013)	2.0	0.7–3.5	3.3	1.4–5.4	3.0	1.2–5.0	3.6	1.6–5.9	0.0075	0.0034–0.0128
Combined data	4.2	1.9–6.6	3.5	1.5–5.6	3.3	1.3–5.6	4.3	1.9–6.6	0.0087	0.0042–0.0145
<i>Entire mitochondrial genome</i>										
Archer et al. (2013)	0.45	0.25–0.68	1.9	1.1–2.8	0.87	0.50–1.3	1.9	1.1–2.8	0.003	0.0018–0.0044
Combined data	0.99	0.54–1.4	1.9	1.1–2.8	0.87	0.48–1.3	1.9	1.1–2.8	0.003	0.0018–0.0044

Notes: TMRCA: the time to the most recent common ancestor, 95% HPD: 95% interval of the highest posterior density. Times are in million years, and the substitution rate is in substitution per site per million years.

\* North Atlantic excluding Mediterranean Sea.

\*\* North Pacific excluding Sea of Cortez.

**Table 3**Average estimates of genetic diversity ( $\theta$ ) and number of immigrants per generation ( $N_e m$ ) for the North Atlantic, North Pacific and Southern Hemisphere\*

Parameter	$\theta_{NA}$	$\theta_{NP}$	$\theta_{SH}$	$N_e m_{NP \rightarrow NA}$	$N_e m_{SH \rightarrow NA}$	$N_e m_{NA \rightarrow NP}$	$N_e m_{SH \rightarrow NP}$	$N_e m_{NA \rightarrow SH}$	$N_e m_{NP \rightarrow SH}$
mode	0.037	0.018	0.078	0.0029	0.3711	0.0015	0.3565	0.0063	0.0063
95% CI	0.022–0.057	0.008–0.032	0.048–0.139	0–1.973	0–2.791	0–1.566	0–3.406	0–3.082	0–18.632

Notes: NA: North Atlantic, NP: North Pacific, SH: Southern Hemisphere,  $\theta$ : genetic diversity,  $N_e$ : effective population size,  $m$ : immigration rate per generation,  $\rightarrow$  denotes the direction of migration, 95% CI: 95% credible interval. \*Estimates were based on the mtDNA CR sequences. Samples from Sea of Cortez and the Mediterranean Sea were excluded from the analysis.

**Table 4**

Multi-locus microsatellite genotype probability value (p-value) per putative population and assigned population.

Sample ID*	CR haplotype number	North Atlantic (n = 266)	Antarctica (n = 20)	Eastern North Pacific (n = 24)	Sea of Cortez (n = 45)	# loci	Missing loci	Assigned population
NAT0009	NATL_011	0.558	< 0.01	< 0.01	< 0.01	20		North Atlantic
NAT0017	NATL_011	0.400	0.045	< 0.01	< 0.01	20		North Atlantic
NAT0019	NATL_011	0.938	0.062	0.049	< 0.01	20		North Atlantic
NAT0024	NATL_011	0.740	< 0.01	< 0.01	< 0.01	20		North Atlantic
NAT0647	NATL_011	0.938	0.062	0.049	< 0.01	20		North Atlantic
NAT0648	NATL_003	0.502	0.025	0.013	< 0.01	19	GT023	North Atlantic
NAT0001	NATL_011	0.547	0.032	0.018	< 0.01	20		North Atlantic
NAT0002	NATL_011	0.692	0.011	< 0.01	< 0.01	20		North Atlantic
NAT0662	NATL_011	0.547	0.032	0.018	< 0.01	20		North Atlantic
NAT0003	NATL_011	0.030	< 0.01	< 0.01	< 0.01	20		North Atlantic
NAT0705	NATL_011	0.394	< 0.01	< 0.01	< 0.01	20		North Atlantic
NAT0004	NATL_011	0.125	0.053	0.019	< 0.01	20		North Atlantic
NAT0706	NATL_011	0.712	0.079	0.060	< 0.01	20		North Atlantic
NAT0707	NATL_016	0.064	< 0.01	< 0.01	< 0.01	20		North Atlantic
NAT0708	NATL_011	0.610	0.012	0.055	< 0.01	20		North Atlantic
NAT0005	NATL_012	0.339	< 0.01	< 0.01	< 0.01	20		North Atlantic
NAT0709	NATL_011	0.223	< 0.01	< 0.01	< 0.01	20		North Atlantic
NAT0276	NATL_011	0.049	< 0.01	< 0.01	< 0.01	20		North Atlantic
NAT0710	NATL_011	0.073	< 0.01	< 0.01	< 0.01	20		North Atlantic
NAT0296	NATL_011	0.213	< 0.01	< 0.01	< 0.01	20		North Atlantic
NAT0711	NATL_011	0.016	< 0.01	< 0.01	< 0.01	20		North Atlantic
NAT0712	NATL_011	0.618	0.055	0.014	< 0.01	20		North Atlantic
SHE0010	SHEM_006	< 0.01	0.190	< 0.01	< 0.01	19	EV001	Antarctic
NPA0347	NPAC_009	< 0.01	0.068	0.938	< 0.01	19	GATA91083	Eastern North Pacific
SOC0172	NPAC_005	< 0.01	< 0.01	0.076	0.169	20		Sea of Cortez

\* The first three letters from the sample ID represent the region where the samples were collected, NAT denotes the North Atlantic Ocean, SHE denotes the Southern Hemisphere, i.e., Antarctic, NPA denotes the Eastern North Pacific Ocean and SOC denotes the Sea of Cortez. The n value in parenthesis represents the sample size for each putative source population. The assigned population was based on the most likely population and its relative score. CR: mtDNA control region.

emergence of polyphyly due to an increased North Atlantic sample size suggests that similar undetected polyphyly may emerge in other ocean basins and regions and hence any inferences made regarding monophyly in these other regions may change as well. This general fundamental sampling issue is problematic in term of defining subspecies from mitogenomic data since the “distinctiveness” may change with the sampling effort. Consequently, such “higher level” intra-specific classifications should not be based solely on uniparentally-inherited, non-recombining genomes. Perhaps employing measures of evolutionary distinctiveness that do not rely upon the “absence” of contradicting observations, e.g., the absence of poly- or paraphyly. Possible, and likely more robust criteria would include the level of gene flow and divergence time or a combination hereof (Hey and Nielsen, 2004; Jackson et al., 2014) inferred from biparentally inherited recombining loci in conjunction with heritable, non-molecular traits, such as ecology and morphology (Crandall et al., 2000). However, settling on exact quantitative criteria for vaguely defined entities, such as, subspecies and ESUs is no simple matter.

The non-trivial nature of defining species and sub-species is, in many instances, hampering implementation of legal protective measures. In response to these practical implications, Taylor et al. (2017) recently proposed employing population genetic statistics (specifically nucleotide divergence) as a means to delineate sub-species in cetaceans. In principle such objective quantitative genetic criteria are desirable, but most many of these statistics are subject to the same sampling and

“rate-of-divergence” issues as a monophyly criterion as discussed above.

Rare, but occasional, gene flow between baleen whale populations from different hemispheres is possible and appeared to have occurred in humpback whales (Jackson et al., 2014) and Antarctic minke whales, *Balaenoptera bonaerensis* (Glover et al., 2010). Estimates of inter-ocean migration rate among fin whales in this study (Table 3) suggested some gene flow between the Southern Hemisphere and the two Northern Hemisphere ocean basins (i.e., the North Atlantic and the North Pacific). However, the wide 95% credible intervals prevented the exclusion of zero migration. The most likely route for recent inter-oceanic gene flow between Northern Hemisphere fin whales populations was through the Southern Hemisphere, i.e., in a stepping stone manner. Such recent, or low ongoing, migration between the Southern Hemisphere and both the North Atlantic and North Pacific, respectively, could explain why some North Pacific and North Atlantic fin whale mtDNA haplotypes clustered within the clade that contained most Southern Hemisphere haplotypes.

The current, accepted classification assigns fin whales from the Northern and Southern Hemisphere to two different subspecies. This taxonomic division implies that North Pacific and North Atlantic fin whales belong to the same subspecies and the Southern Hemisphere fin whales to another subspecies. This taxonomic classification was based upon differences in the vertebrae as well as size differences (Lönnberg, 1931; Tomilin, 1946). The basis of these differences has since been

questioned by Perrin et al. (2009), who suggested that the different latitudinal origin of the holotypes might explain the observed size difference (Perrin et al., 2009). However, this explanation is difficult to evaluate since the holotype that served as the basis for the differences in the vertebrae described by Lönnberg (1931) was not collected and hence is unavailable. Alternatively, if the Northern Hemisphere populations were founded from the Southern Hemisphere the observed polyphyly could be due to incomplete lineage sorting (Avice et al., 1984) as suggested by Pastene et al. (2007) in the case of common minke whales, *Balaenoptera acutorostrata*, in the Atlantic Ocean. This appears to be the inference drawn by Archer et al. (2013), who emphasizes that the three well supported North Pacific groups (Fig. 3a) observed in their mitogenome-based genealogy could be due to incomplete lineage sorting. However, whether the patterns observed in the taxon (i.e., monophyletic, polyphyletic or paraphyletic groups) represent sub-species, as opposed to incomplete lineage sorting, population structure and/or incomplete sampling remains unclear.

Unsurprisingly no qualitative differences between the topologies inferred for the basal part of the mitochondrial genealogies were detected when increasing the data from only 285 bp of mtDNA CR sequence to the complete mitogenome DNA sequences. The general support for individual nodes, especially the most recent nodes, increased with the number of base pairs per haplotype and hence node support was substantially higher in genealogy estimated based upon complete mitogenome haplotypes. However, in most cases, the basic nodes are the target of interest in analyses of intraspecific variation aimed at assessing sub-species or ESUs. This observation, together with the obvious need for an increased sampling coverage, suggests that it might be worthwhile to first sequence a limited number of mitogenomes from the extreme parts of the species' distribution. The mitogenome sequences can then serve as a backbone to identify and subsequently specifically target informative regions, which likely can be sequenced efficiently and at low costs using "standard" Sanger sequencing as proposed by Coulson et al. (2006). Such a strategy, as opposed to next-generation sequencing of the entire mitogenome in all specimens, would facilitate large sample sizes presumably with minimal loss of phylogenetic signal in terms the basic nodes.

In conclusion, the present study found that some of the spatially distinct mtDNA haplotype monophyly in North Atlantic fin whales reported by Archer et al. (2013) was due to an insufficient sample size. Although untested in this study, the same could be the case for some of the monophyly detected in other ocean basins or regions. Since monophyly essentially relies upon "absence of evidence" for poly or paraphyly proving monophyly, especially below the species level is difficult and prone to biases. As pointed out by Crandall et al., 2000 identifying sub-species or ESUs solely from genetic data is an oversimplification and should be complemented with ecological, behavioural and morphological data. In principle, genetic data are well-suited to assess divergence times and the degree of reproductive isolation (when gene flow is low) but the choice of suitable statistics and appropriate threshold values is no simple task.

## Acknowledgements

We are grateful to Hanne Jørgensen, Anna Sellas, Mary Beth Rew and Christina Færch-Jensen for technical assistance. We thank Drs. P. E. Rosel and K. D. Mullin (U.S. National Marine Fisheries Service Southeast Fisheries Science Center) and members of the U.S. Northeast and Southeast Region Marine Mammal Stranding Network and its response teams, including the International Fund for Animal Welfare, the Marine Mammal Stranding Center, Mystic Aquarium, the Riverhead Foundation for Marine Research and Preservation (K. Durham) and the Marine Mammal Stranding Program of the University of North Carolina Wilmington for access to fin whale samples from the western North Atlantic. We thank Gisli Víkingsson for providing samples. We are indebted to Dr. Eduardo Secchi for facilitating data sharing. Data

collection in the Southern Ocean was conducted under research projects Baleias (CNPq grants 557064/2009-0 and 408096/2013-6), INTERBIOTA (CNPq 407889/2013-2) and INCT-APA (CNPq 574018/2008-5), of the Brazilian Antarctic Program and a contribution by the research consortium 'Ecology and Conservation of Marine Megafauna – EcoMega-CNPq'. MAS was supported through a FCT Investigator contract funded by POPH, QREN European Social Fund, and Portuguese Ministry for Science and Education (IF/00943/2013). RP was supported by an FCT postdoctoral grant (SFRH/BPD/108007/2015). We acknowledge funds provided by FCT to MARE, through the strategic project UID/MAR/04292/2019. Data collection in the Azores was funded by TRACE-PTDC/MAR/74071/2006 and MAPCET-M2.1.2/F/012/2011 [FEDER, COMPETE, QREN European Social Fund, and Proconvergencia Açores/EU Program]. AAC was supported by University of Groningen. VR-L was funded by Consejo Nacional de Ciencia y Tecnología (CONACYT). Fin whale illustration herein is used with the permission of Frédérique Lucas. We acknowledge the Center for Information Technology of the University of Groningen for IT support and access to the Peregrine high performance-computing cluster.

## Contributions to the paper

PJP, MB, JPAH and AAC conceived and designed the study. AA, SGB, SB, DB, AB, HAC, LDR, PG, SL, FL, VM, SM, NO, CP, SP, RP, CR, JR, CR, RS, MAS, JU, GV, FWW provided data or sample material, MB, JPAH, CPD, WH, VR-L conducted laboratory analyses. AAC conducted the data analysis with contributions from JPAH and inputs from ES. TO conducted the Web of Science review. AAC, JPAH, PJP and MB drafted the manuscript. All authors read, edited, commented on and approved the final manuscript.

## Declaration of interest

None.

## Data accessibility

Mitochondrial DNA sequence data, microsatellite genotypes and input data files for the analyses conducted in this manuscript have been deposited in Datadryad.org under accession: <https://doi.org/10.5061/dryad.qt528n0>.

## Appendix A. Supplementary material

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.ympev.2019.02.003>.

## References

- Amos, B., Hoelzel, A.R., 1991. Long-term preservation of whale skin for DNA analysis. *Rep. Int. Whaling Commission, Special Issue 13*, 99–104.
- Archer, F.I., Morin, P.A., Hancock-Hanser, B.L., Robertson, K.M., Leslie, M.S., Bérubé, M., Taylor, B.L., 2013. Mitogenomic phylogenetics of fin whales (*Balaenoptera physalus* spp.): genetic evidence for revision of subspecies. *PLoS ONE* 8 (5), e63396. <https://doi.org/10.1371/journal.pone.0063396>.
- Árnason, U., Gullberg, A., Widegren, B., 1991. The complete nucleotide sequence of the mitochondrial DNA of the fin whale, *Balaenoptera physalus*. *J. Mol. Evol.* 33 (6), 556–568. <https://doi.org/10.1007/bf02102808>.
- Avice, J.C., 1989. Gene trees and organismal histories: a phylogenetic approach to population biology. *Evolution* 43 (6), 1192–1208. <https://doi.org/10.2307/2409356>.
- Avice, J.C., Arnold, J., Ball, R.M., Bermingham, E., Lamb, T., Neigel, J.E., Saunders, N.C., 1987. Intraspecific phylogeography: the mitochondrial DNA bridge between population genetics and systematics. *Annu. Rev. Ecol. Syst.* 18, 489–522. <https://doi.org/10.1146/annurev.ecolsys.18.1.489>.
- Avice, J.C., Giblin-Davidson, C., Laerm, J., Patton, J.C., Lansman, R.A., 1979. Mitochondrial DNA clones and matriarchal phylogeny within and among geographic populations of the pocket gopher, *Geomys pinetis*. *PNAS* 76 (12), 6694–6698. <https://doi.org/10.1073/pnas.76.12.6694>.
- Avice, J.C., Neigel, J.E., Arnold, J., 1984. Demographic influences on mitochondrial DNA lineage survivorship in animal populations. *J. Mol. Evol.* 20 (2), 99–105. <https://doi.org/10.1007/Bf02257369>.



- Ball, R.M., Avise, J.C., 1992. Mitochondrial DNA phylogeographic differentiation among avian populations and the evolutionary significance of subspecies. *Auk* 109 (3), 626–636.
- Banguera-Hinestroza, E., Cárdenas, H., Ruiz-García, M., Marmontel, M., Gaitán, E., Vázquez, R., García-Vallejo, F., 2002. Molecular identification of evolutionarily significant units in the Amazon River Dolphin *Inia sp.* (Cetacea:Iniidae). *J. Hered.* 93 (5), 312–322.
- Beerli, P., Felsenstein, J., 1999. Maximum-likelihood estimation of migration rates and effective population numbers in two populations using a coalescent approach. *Genetics* 152 (2), 763–773.
- Beerli, P., Felsenstein, J., 2001. Maximum likelihood estimation of a migration matrix and effective population sizes in *n* subpopulations by using a coalescent approach. *Proc. Nat. Acad. Sci. United States of America* 98 (8), 4563–4568. <https://doi.org/10.1073/pnas.081068098>.
- Bernatchez, L., 1995. A role for molecular systematics in defining evolutionarily significant units in fishes. In: Nielsen, J.L. (Ed.), *Evolution and the Aquatic Ecosystem. Defining Unique Units in Population Conservation*. American Fisheries Society Symposium, Bethesda, Maryland, pp. 114–132.
- Bérubé, M., Aguilar, A., Dendanto, D., Larsen, F., Notarbartolo Di Sciarra, G., Sears, R., Palsbøll, P.J., 1998. Population genetic structure of North Atlantic, Mediterranean Sea and Sea of Cortez fin whales, *Balaenoptera physalus* (Linnaeus 1758): analysis of mitochondrial and nuclear loci. *Mol. Ecol.* 7 (5), 585–599. <https://doi.org/10.1046/j.1365-294x.1998.00359.x>.
- Bérubé, M., Jørgensen, H., McEwing, R., Palsbøll, P.J., 2000. Polymorphic di-nucleotide microsatellite loci isolated from the humpback whale, *Megaptera novaeangliae*. *Molecular Ecology* 9 (12), 2181–2183. <https://doi.org/10.1046/j.1365-294X.2000.105315.x>.
- Bérubé, M., Palsbøll, P., 1996a. Erratum of identification of sex in cetaceans by multiplexing with three ZFX and ZFY specific primers. *Mol. Ecol.* 5 (4), 602. <https://doi.org/10.1111/j.1365-294X.1996.tb00355.x>.
- Bérubé, M., Palsbøll, P., 1996b. Identification of sex in cetaceans by multiplexing with three ZFX and ZFY specific primers. *Mol. Ecol.* 5 (2), 283–287.
- Bérubé, M., Rew, M., Skaug, H., Jørgensen, H., Robbins, J., Best, P., Palsbøll, P., 2005. Polymorphic microsatellite loci isolated from humpback whale, *Megaptera novaeangliae* and fin whale, *Balaenoptera physalus*. *Conserv. Genet.* 6 (4), 631–636. <https://doi.org/10.1007/s10592-005-9017-5>.
- Bérubé, M., Urban, J., Dizon, A.E., Brownell, R.L., Palsbøll, P.J., 2002. Genetic identification of a small and highly isolated population of fin whales (*Balaenoptera physalus*) in the Sea of Cortez, Mexico. *Conserv. Genet.* 3 (2), 183–190. <https://doi.org/10.1023/a:1015224730394>.
- Burbink, F.T., Lawson, R., Slowinski, J.B., 2000. Mitochondrial DNA phylogeography of the polytypic North American rat snake (*Elaphe obsoleta*): a critique of the subspecies concept. *Evolution* 54 (6), 2107–2118. <https://doi.org/10.1111/j.0014-3820.2000.tb01253.x>.
- Coates, A.G., Jackson, J.B.C., Collins, L.S., Cronin, T.M., Dowsett, H.J., Bybell, L.M., Obando, J.A., 1992. Closure of the Isthmus of Panama: the near-shore marine record of Costa Rica and Western Panama. *Geol. Soc. Am. Bull.* 104 (7), 814–828. <https://doi.org/10.1130/0016-7606>.
- Coulson, M.W., Marshall, H.D., Pepin, P., Carr, S.M., 2006. Mitochondrial genomics of gadine fishes: implications for taxonomy and biogeographic origins from whole-genome data sets. *Genome* 49 (9), 1115–1130. <https://doi.org/10.1139/G06-083>.
- Crandall, K.A., Bininda-Emonds, O.R.P., Mace, G.M., Wayne, R.K., 2000. Considering evolutionary processes in conservation biology. *Trends Ecol. Evol.* 15 (7), 290–295. [https://doi.org/10.1016/S0169-5347\(00\)01876-0](https://doi.org/10.1016/S0169-5347(00)01876-0).
- Darriba, D., Taboada, G.L., Doallo, R., Posada, D., 2012. jModelTest 2: more models, new heuristics and parallel computing. *Nat. Methods* 9 (8), 772.
- Davis, R.W., Fargion, G.S., May, N., Leming, T.D., Baumgartner, M., Evans, W.E., Hansen, L.J., Mullin, K., 1998. Physical habitat of cetaceans along the continental shelf in the North-Central and Western Gulf of Mexico. *Marine Mammal Sci.* 14 (3), 490–507.
- Dawbin, W.H., 1966. The seasonal migratory cycle of humpback whales. In: Norris, K.S. (Ed.), *Whales, dolphins and porpoises*. University of California Press, Berkeley, CA, pp. 145–170.
- Drouot, V., Berube, M., Gannier, A., Goold, J.C., Reid, R.J., Palsbøll, P.J., 2004. A note on genetic isolation of Mediterranean sperm whales (*Physeter macrocephalus*) suggested by mitochondrial DNA. *J. Cetacean Res. Manage.* 6 (1), 29–32.
- Drummond, A.J., Rambaut, A., 2007. BEAST: Bayesian evolutionary analysis by sampling trees. *BMC Evol. Biol.* 7 (1), 214. <https://doi.org/10.1186/1471-2148-7-214>.
- Drummond, A.J., Suchard, M.A., Xie, D., Rambaut, A., 2012. Bayesian Phylogenetics with BEAUti and the BEAST 1.7. *Mol. Biol. Evol.* 29 (8), 1969–1973. <https://doi.org/10.1093/molbev/mss075>.
- Funk, D.J., Omland, K.E., 2003. Species-level paraphyly and polyphyly: Frequency, causes, and consequences, with insights from animal mitochondrial DNA. *Annu. Rev. Ecol. Syst.* 34, 397–423. <https://doi.org/10.1146/annurev.ecolsys.34.011802.132421>.
- Funk, W.C., McKay, J.K., Hohenlohe, P.A., Allendorf, F.W., 2012. Harnessing genomics for delineating conservation units. *Trends Ecol. Evol.* 27 (9), 489–496. <https://doi.org/10.1016/j.tree.2012.05.012>.
- Gambell, R., 1985. Fin whale *Balaenoptera physalus* (Linnaeus, 1758). In: Ridgway, S.H., Harrison, S.R. (Eds.), *Handbook of Marine Mammals*. Academic Press Inc, London, UK, pp. 171–192.
- Glover, K.A., Kanda, N., Haug, T., Pastene, L.A., Øien, N., Goto, M., Seliussen, B.B., Skaug, H.J., 2010. Migration of Antarctic minke whales to the Arctic. *PLoS One* 5 (12). <https://doi.org/10.1371/journal.pone.0015197>.
- Halbert, K.M.K., Goetze, E., Carlson, D.B., 2013. High cryptic diversity across the global range of the migratory planktonic copepods *Pleuromamma pisces* and *P. gracilis*. *Plos One* 8 (10). <https://doi.org/10.1371/journal.pone.0077011>.
- Hasegawa, M., Kishino, H., Yano, T.A., 1985. Dating of the human – ape splitting by a molecular clock of mitochondrial DNA. *J. Mol. Evol.* 22 (2), 160–174. <https://doi.org/10.1007/Bf02101694>.
- Hey, J., Nielsen, R., 2004. Multilocus methods for estimating population sizes, migration rates and divergence time, with applications to the divergence of *Drosophila pseudoobscura* and *D. persimilis*. *Genetics* 167 (2), 747–760. <https://doi.org/10.1534/genetics.103.024182>.
- Hudson, R.R., Turelli, M., 2003. Stochasticity overrules the “three-times rule”: genetic drift, genetic draft, and coalescence times for nuclear loci versus mitochondrial DNA. *Evolution* 57 (1), 182–190. <https://doi.org/10.1111/j.0014-3820.2003.tb00229.x>.
- Ingebrigtsen, A., 1929. Whales caught in the North Atlantic and other seas. Conseil Permanent International pour l'Exploration de la Mer. Rapports et Procès-Verbaux des Reunions 56, 123–135.
- Jackson, J.A., Steel, D.J., Beerli, P., Congdon, B.C., Olavarria, C., Leslie, M.S., Pomilla, C., Rosenbaum, H., Baker, C.S., 2014. Global diversity and oceanic divergence of humpback whales (*Megaptera novaeangliae*). *Proc. Royal Soc. London, Series B: Biol. Sci.* 281 (1786). <https://doi.org/10.1098/rspb.2013.3222>.
- Jonggård, Å., 1966. The distribution of Balaenopteridae in the north Atlantic Ocean. In: Norris, K.S. (Ed.), *Whales, dolphins and porpoises*. University of California Press, Berkeley, CA, pp. 114–124.
- Katona, S.K., Whitehead, H.P., 1981. Identifying humpback whales using their natural markings. *Polar Res.* 20 (128), 439–444.
- Leaché, A.D., 2009. Species tree discordance traces to phylogeographic clade boundaries in North American fence lizards (*Sceloporus*). *Syst. Biol.* 58 (6), 547–559. <https://doi.org/10.1093/sysbio/syp057>.
- Lönnberg, E., 1931. The skeleton of *Balaenoptera brydei* Ö. Almqvist & Wiksell, Olsen.
- Meng, X.P., Shen, X., Zhao, N.N., Tian, M., Liang, M., Hao, J., Zhu, X.L., 2013. Mitogenomics reveals two subspecies in *Coelomacra antiquata* (Mollusca: Bivalvia). *Mitochondrial DNA* 24 (2), 102–104. <https://doi.org/10.3109/19401736.2012.726620>.
- Morin, P.A., Archer, F.I., Foote, A.D., Vilstrup, J., Allen, E.E., Wade, P., Harkins, T., 2010. Complete mitochondrial genome phylogeographic analysis of killer whales (*Orcinus orca*) indicates multiple species. *Genome Res.* 20 (7), 908–916.
- Morin, P.A., Luikart, G., Wayne, R.K., Grp, S.W., 2004. SNPs in ecology, evolution and conservation. *Trends Ecol. Evol.* 19 (4), 208–216. <https://doi.org/10.1016/j.tree.2004.01.009>.
- Moritz, C., 1994. Defining 'Evolutionarily Significant Units' for conservation. *Trends Ecol. Evol.* 9 (10), 373–375. [https://doi.org/10.1016/0169-5347\(94\)90057-4](https://doi.org/10.1016/0169-5347(94)90057-4).
- Mullis, K.B., Faloona, F.A., 1987. Specific synthesis of DNA in vitro via a polymerase-catalyzed chain reaction. *Methods Enzymol.* 155, 335–350. [https://doi.org/10.1016/0076-6879\(87\)55023-6](https://doi.org/10.1016/0076-6879(87)55023-6).
- Paetkau, D., 1999. Using genetics to identify intraspecific conservation units: a critique of current methods. *Conserv. Biol.* 13 (6), 1507–1509. <https://doi.org/10.1046/j.1523-1739.1999.98507.x>.
- Paetkau, D., Strobeck, C., 1994. Microsatellite analysis of genetic variation in black bear populations. *Mol. Ecol.* 3 (5), 489–495. <https://doi.org/10.1111/j.1365-294X.1994.tb00127.x>.
- Page, R.D.M., Charleston, M.A., 1997. From gene to organismal phylogeny: Reconciled trees and the gene tree species tree problem. *Mol. Phylogenet. Evol.* 7 (2), 231–240. <https://doi.org/10.1006/mpev.1996.0390>.
- Palsbøll, P.J., Bérubé, M., Aguilar, A., Notarbartolo-Di-Sciarra, G., Nielsen, R., 2004. Discerning between recurrent gene flow and recent divergence under a finite-site mutation model applied to North Atlantic and Mediterranean Sea fin whale (*Balaenoptera physalus*) populations. *Evolution* 58 (3), 670–675. <https://doi.org/10.1111/j.0014-3820.2004.tb01691.x>.
- Palsbøll, P.J., Bérubé, M., Larsen, A.H., Jørgensen, H., 1997. Primers for the amplification of tri- and tetramer microsatellite loci in baleen whales. *Mol. Ecol.* 6 (9), 893–895. <https://doi.org/10.1046/j.1365-294X.1997.d01-214.x>.
- Palsbøll, P.J., Clapham, P.J., Mattila, D.K., Larsen, F., Sears, R., Siegmund, H.R., Sigurjónsson, J., Vasquez, O., Arctander, P., 1995. Distribution of mtDNA haplotypes in North Atlantic humpback whales: the influence of behaviour on population structure. *Marine Ecol. Progress Series* 116, 1–10.
- Palsbøll, P.J., Larsen, F., Hansen, E.S., 1991. Sampling of skin biopsies from free-ranging large cetaceans in West Greenland: Development of new biopsy tips and bolt designs. Report of the International Whaling Commission (Special Issue 13) 71–79.
- Palumbi, S.R., Baker, C.S., 1994. Contrasting population structure from nuclear intron sequences and mtDNA of humpback whales. *Mol. Biol. Evol.* 11 (3), 426–435.
- Pamilo, P., Nei, M., 1988. Relationships between gene trees and species trees. *Mol. Biol. Evol.* 5 (5), 568–583.
- Pastene, L.A., Goto, M., Kanda, N., Zerbini, A.N., Kerem, D., Watanabe, K., Palsbøll, P.J., 2007. Radiation and speciation of pelagic organisms during periods of global warming: the case of the common minke whale, *Balaenoptera acutorostrata*. *Molecular Ecol.* 16 (7), 1481–1495. <https://doi.org/10.1111/j.1365-294X.2007.03244.x>.
- Perrin, W.F., Mead, J.G., Brownell Jr, R.L., 2009. Review of the evidence used in the description of currently recognized cetacean subspecies NOAA Technical Memorandum NMFS - SWFSC-450.
- Piry, S., Alapetite, A., Cornuet, J.M., Paetkau, D., Baudouin, L., Estoup, A., 2004. GENECLASS2: a software for genetic assignment and first-generation migrant detection. *J. Hered.* 95 (6), 536–539. <https://doi.org/10.1093/jhered/esh074>.
- Plummer, M., Best, N., Cowles, K., Vines, K., 2006. CODA: Convergence diagnosis and output analysis for MCMC. *R News* 6, 7–11.
- Pons, J., Barraclough, T.G., Gomez-Zurita, J., Cardoso, A., Duran, D.P., Hazell, S., Vogler, A.P., 2006. Sequence-based species delimitation for the DNA taxonomy of undescribed insects. *Syst. Biol.* 55 (4), 595–609. <https://doi.org/10.1080/10635150600852011>.
- Prager, E.M., Sage, R.D., Gyllenstein, U., Thomas, W.K., Hubner, R., Jones, C.S., Wilson,



- A.C., 1993. Mitochondrial DNA sequence diversity and the colonization of Scandinavia by house mice from East Holsten. *Biol. J. Linn. Soc.* 50 (2), 85–122.
- Rambaut, A., Drummond, A.J., 2007. Tracer v1. 4.
- Rice, C.W., 1998. Marine mammals of the world, systematics and distribution: Society for Marine Mammalogy Special Publications 4.
- Rosenbaum, H.C., Brownell, R.L., Brown, M.W., Schaeff, C., Portway, V., White, B.N., DeSalle, R., 2000. World-wide genetic differentiation of *Eubalaena*: questioning the number of right whale species. *Mol. Ecol.* 9 (11), 1793–1802. <https://doi.org/10.1046/j.1365-294x.2000.01066.x>.
- Ryder, O.A., 1986. Species conservation and systematics: the dilemma of subspecies. *Trends Ecol. Evol.* 1 (1), 9–10. [https://doi.org/10.1016/0169-5347\(86\)90059-5](https://doi.org/10.1016/0169-5347(86)90059-5).
- Sambrook, J., Russell, D.W., 2001. *Molecular Cloning: a Laboratory Manual, Third ed.* Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, USA.
- Sanger, F., 1981. Determination of nucleotide sequences in DNA. *Science* 214 (4526), 1205–1210. <https://doi.org/10.1126/science.7302589>.
- Sasaki, T., Nikaido, M., Hamilton, H., Goto, M., Kato, H., Kanda, N., Okada, N., 2005. Mitochondrial phylogenetics and evolution of mysticete whales. *Syst. Biol.* 54 (1), 77–90. <https://doi.org/10.1080/10635150590905939>.
- Taylor, B.L., Archer, F.I., Martien, K.K., Rosel, P.E., Hancock-Hanser, B.L., Lang, A.R., Baker, C.S., 2017. Guidelines and quantitative standards to improve consistency in cetacean subspecies and species delimitation relying on molecular genetic data. *Mar. Mammal Sci.* 33, 132–155. <https://doi.org/10.1111/mms.12411>.
- Tomilin, A.G., 1946. Thermoregulation and the geographical races of cetaceans (Termoregulyatsiya i geograficheskie racy kitoobraznykh.). *Doklady Akad. Nauk CCCP* 54, 465–472.
- Valsecchi, E., Amos, W., 1996. Microsatellite markers for the study of cetacean populations. *Mol. Ecol.* 5 (1), 151–156. <https://doi.org/10.1111/j.1365-294X.1996.tb00301.x>.
- Valsecchi, E., Palsbøll, P., Hale, P., GlocknerFerrari, D., Ferrari, M., Clapham, P., Amos, B., 1997. Microsatellite genetic distances between oceanic populations of the humpback whale (*Megaptera novaeangliae*). [Article]. *Mol. Biol. Evol.* 14 (4), 355–362.
- Vilstrup, J.T., Ho, S.Y., Foote, A.D., Morin, P.A., Kreb, D., Krützen, M., Parra, G.J., Robertson, K.M., de Stephanis, R., Verborgh, P., Willerslev, E., Orlando, L., Gilbert, M.T.P., 2011. Mitogenomic phylogenetic analyses of the Delphinidae with an emphasis on the Globicephalinae. *BMC Evol. Biol.* 11 (65). <https://doi.org/10.1186/1471-2148-11-65>.
- Zachos, F.E., Apollonio, M., Barmann, E.V., Festa-Bianchet, M., Gohlich, U., Habel, J.C., Suchentrunk, F., 2013. Species inflation and taxonomic artefacts-A critical comment on recent trends in mammalian classification. *Mamm. Biol.* 78 (1), 1–6. <https://doi.org/10.1016/j.mambio.2012.07.083>.